

Apolipoprotein B genetic variants modify the response to fenofibrate: a GOLDN study^S

Mary K. Wojczynski,* Guimin Gao,* Ingrid Borecki,[†] Paul N. Hopkins,[§] Laurence Parnell,** Chao-Qiang Lai,** Jose M. Ordovas,** B. Hong Chung,^{††} and Donna K. Arnett^{1,§§}

Section on Statistical Genetics,* Department of Biostatistics, University of Alabama at Birmingham, Birmingham, AL; Division of Statistical Genomics,[†] Washington University School of Medicine, St. Louis, MO; Cardiovascular Genetics,[§] Cardiology Division, University of Utah, Salt Lake City, UT; Nutrition and Genomics Laboratory,** Jean Mayer-US Department of Agriculture Human Nutrition Research Center on Aging, Tufts University, Boston, MA; Department of Nutrition Sciences,^{††} School of Health Professions, University of Alabama at Birmingham, AL; and Department of Epidemiology,^{§§} School of Public Health and Clinical Nutrition Research Center, University of Alabama at Birmingham, Birmingham, AL

Abstract Hypertriglyceridemia, defined as a triglyceride measurement > 150 mg/dl, occurs in up to 34% of adults. Fenofibrate is a commonly used drug to treat hypertriglyceridemia, but response to fenofibrate varies considerably among individuals. We sought to determine if genetic variation in apolipoprotein B (APOB), an essential core of triglyceride-rich lipoprotein formation, may account for some of the inter-individual differences observed in triglyceride (TG) response to fenofibrate treatment. Participants (N = 958) from the Genetics of Lipid Lowering Drugs and Diet Network study completed a three-week intervention with fenofibrate 160 mg/day. Associations of four APOB gene single nucleotide polymorphisms (SNP) (rs934197, rs693, rs676210, and rs1042031) were tested for association with the TG response to fenofibrate using a mixed growth curve model where the familial structure was modeled as a random effect and cardiovascular risk factors were included as covariates. Three of these four SNPs changed the amino acid sequence of APOB, and the fourth was in the promoter region. TG response to fenofibrate treatment was associated with one APOB SNP, rs676210 (Pro2739Leu), such that participants with the TT genotype of rs676210 had greater TG lowering than those with the CC genotype (additive model, $P = 0.0017$).^{¶¶} We conclude the rs676210 variant may identify individuals who respond best to fenofibrate for TG reduction.—Wojczynski, M. K., G. Gao, I. Borecki, P. N. Hopkins, L. Parnell, C-Q. Lai, J. M. Ordovas, B. H. Chung, and D. K. Arnett. **Apolipoprotein B genetic variants modify**

the response to fenofibrate: a GOLDN study. *J. Lipid Res.* 2010. 51: 3316–3323.

Supplementary key words fenofibrate • polymorphism • triglycerides • Genetics of Lipid Lowering Drugs and Diet Network

Hypertriglyceridemia and mixed dyslipidemia [i.e., high levels of low-density lipoprotein cholesterol (LDL-C) and triglyceride (TG) levels combined with decreased levels of high-density lipoprotein cholesterol (HDL-C)] contribute to the development and progression of atherosclerosis (1, 2). Plasma lipid and apolipoprotein levels are influenced by both genetic variation and environmental factors, particularly diet, exercise, and smoking (3, 4). Hypertriglyceridemia, defined as TG > 150 mg/dl, is a common condition, occurring in approximately one-third of adults, and in the United States about 1.3% of the population uses fenofibrate, gemfibrozil, or niacin to lower triglyceride (5). Treatment with fibrates primarily reduces TG (21–35%), but it also reduces LDL-C (9–11%) and increases HDL-C (5–18%) (6). Fenofibrate, a derivative of fibric acid, is an oral pharmacologic agent that is prescribed for mixed dyslipidemia and hypertriglyceridemia in adults that are nonresponsive to lifestyle interventions (7). Fenofibrate activates the nuclear transcription factor peroxisome proliferator-activated receptor (PPAR α) in a process that ultimately modulates the expression of genes regulating lipid metabolism. Fenofibrate also enhances β -oxidation of fatty

This work was supported by the Heart, Lung, and Blood Institute, National Institutes of Health, Grant U01 HL-72524 (Genetic and Environmental Determinants of Triglycerides); the Statistical Genetics Post-Doctoral Training Program T32HL07275057, University of Alabama at Birmingham (M.K.W.); National Institutes of Health Grant U54CA100949 (Barnes, PI) at the University of Alabama at Birmingham (G.G.); and the National Institute of General Medical Sciences, National Institutes of Health, Grant R01 GM-073766 (G.G., PI). Its contents are solely the responsibility of the authors and do not necessarily represent the official views of the National Institutes of Health or other granting agencies.

Manuscript received 1 September 2009 and in revised form 19 August 2010.

Published, JLR Papers in Press, August 19, 2010
DOI 10.1194/jlr.P001834

Abbreviations: APOB, apolipoprotein B; PPAR α , peroxisome proliferator-activated receptor α ; SNP, single nucleotide polymorphism; TG, triglyceride.

¹To whom correspondence should be addressed.

e-mail: arnett@uab.edu

^SThe online version of this article (available at <http://www.jlr.org>) contains supplementary data in the form of four tables.

acids, thereby reducing the amount of fatty acids available for TG synthesis (7).

Apolipoproteins are the structural components of lipoprotein particles and serve as cofactors for lipid-metabolizing enzymes and ligands for lipoprotein receptors (8). Apolipoprotein B (APOB) is a requisite protein constituent of LDL, very low-density lipoprotein (VLDL), and chylomicrons (9), transporting both triglyceride and cholesterol esters during lipoprotein metabolism (10). APOB is necessary for the cellular uptake and catabolism of LDL by the LDL receptor. Thus, APOB is critical for the synthesis, transport, and catabolism of TG-rich and cholesterol-rich lipoproteins in the intestine and liver (11, 12). Polymorphisms in the *APOB* gene have been associated with the variability of serum cholesterol levels and coronary atherosclerosis in some populations (13) but not all (14). No studies have examined the role of *APOB* genetic variants in relation to the lipid-lowering effects of fenofibrate.

In this study, we examine the role of four polymorphisms in the *APOB* gene on the response of triglyceride levels to a three-week fenofibrate intervention of 160 mg/day of micronized fenofibrate among individuals participating in the Genetics of Lipid Lowering Drugs and Diet Network (GOLDN) study to further understand the variability in inter-individual response to fenofibrate treatment.

METHODS

Study design

The analyzed study sample included 958 individuals (453 men and 505 women) participating in the GOLDN Study. The GOLDN Study is a component of the PROGRAM for GENetic Interaction (PROGENI) Network, which is a group of family studies examining gene-environment interactions through the use of controlled interventions. This study is funded by the National Institutes of Health (NIH) through the University of Alabama at Birmingham in collaboration with the University of Utah, Washington University, Tufts University, University of Texas, University of Michigan, University of Minnesota, and Fairview-University of Minnesota Medical Center. Participants in the GOLDN Study were recruited from three-generational pedigrees previously identified in the Minneapolis, MN, and Salt Lake City, UT, field centers of the National Heart, Lung, and Blood Institute Family Heart Study (15). The use of these field centers allowed for a genetically homogeneous population (16), as all participants were Caucasian from European ancestry. Eligibility criteria included: age ≥ 18 years; fasting triglycerides < 1500 mg/dl; willingness to participate and attend clinic visits; being a member of a family with at least two members in a sibship; and normal renal and hepatic function (AST and ALT within normal range and creatinine ≤ 2.0 mg/dl). Potential participants were excluded if they reported a history of liver, kidney, pancreas, or gall bladder disease or malabsorption; women currently pregnant; current use of insulin, warfarin, or lipid-lowering drugs [including prescription, over the counter, and nutraceuticals (volunteers taking these agents were withdrawn from their for at least four weeks prior to the study with physician's approval)]; women of childbearing potential not using an acceptable form of contraception; known hypersensitivity to fenofibrate; or history of pancreatitis within 12 months prior to enrollment. Written informed consent was ob-

tained from each participant. The Institutional Review Board at each participating institution approved the study protocol.

The main aim of the GOLDN Study is to characterize the genetic basis of individual variability in response to triglycerides in response to a three-week, open label clinical trial of once daily 160 mg micronized fenofibrate (TriCor®, Abbott Laboratories, Chicago, IL). Three weeks was decided as appropriate for our trial for two reasons: first, a steady-state concentration of fenofibrate is reached at three weeks, and second, a shorter duration of treatment will identify those who are most and least responsive to treatment.

Measurements

We obtained clinical and biochemical measurements of all participating individuals before and after response to fenofibrate. For each clinic visit, participants were asked to abstain from using alcohol for at least 24 h and to fast for at least 12 h. We completed anthropometric measurements, assessed current medications, blood pressure, medical history, dietary history, and personal history.

Blood was also collected to obtain DNA for use in genotyping single nucleotide polymorphisms (SNP) for genetic analyses. Additionally, at visits 1–4, blood was collected for biochemical measurements. Centrifugation of blood samples occurred within 20 min of collection at 2000 *g* for 15 min at 4°C. Collected plasma samples from each participant at each time point were stored at 4°C until study completion. Upon study completion, we analyzed all plasma samples for the same individual at the same time. We made TG measurements using the glycerol-blanked enzymatic method on the Roche COBAS FARA centrifugal analyzer (Roche Diagnostics, Basel, Switzerland). HDL-C measurement used the same procedure after precipitation of nonHDL cholesterol with magnesium/dextran. LDL-C measurement employed a homogeneous direct method (LDL Direct Liquid Select™ Cholesterol Reagent; Equal Diagnostics, Exton, PA) on a Roche Hitachi 911 Automatic Analyzer.

We isolated genomic DNA from peripheral blood leukocytes using the Puregene® reagents following the vendor's protocol. We genotyped five SNPs in the *APOB* gene using a TaqMan assay with allele-specific probes using the ABIPrism 7900HT Sequence Detection System (Applied Biosystems, Foster City, CA) according to standard laboratory protocols (17). Specifically, we analyzed the following SNPs in the *APOB* gene: -516 C to T (rs934197), *XbaI* (rs693), *EcoRI* (rs1042031), Val591Ala (rs679899), and Pro2739Leu (rs676210). SNP selection was performed on phase I HapMap data, as that was all that was available at that time. Selection was based on literature reports of established associations, particularly that of Bentzen et al. (18) for three SNPs (rs679899, rs676210, and rs1042031). Prior results from the lab of Dr. Ordovas (in the context of dietary lipids, not fenofibrate treatment) for rs934197 and rs693 led to inclusion of these SNPs (19–22). Computational analysis of allele-specific predictions of transcription factor binding (PATCH (23)) predicts an allele-specific SP1 site encompassing rs934197.

The overall genotyping success rate for the GOLDN Study was 90%. Approximately 10% of the samples were genotyped twice, once as blind replicates. The agreement among blind replicates assessed using the κ coefficient was ≥ 0.95 for all *APOB* SNPs.

Statistical analysis

TG concentrations were log-transformed to normalize the TG distribution. To determine significant differences in percentages between men or women with and without a known cardiovascular disease risk factor, we used the Pearson χ^2 and Fisher's tests. To compare crude means, we used ANOVA and the Student's *t*-test. We performed statistical analyses using SAS 9.1 (SAS Institute, Inc., Cary, NC).

We used a growth curve mixed model (24, 25) to assess associations between the *APOB* polymorphisms and fenofibrate treatment. We fitted a three-level and individual growth mixed model using SAS Proc Mixed. The first level analyzed individual measurements across two time points for the fenofibrate intervention (pre- and post-treatment with fenofibrate). The second level analyzed the individual nested within pedigree, and the third level examined pedigree effects. To model the individual growth curve, we used an unstructured covariance matrix, treating the intercept and time as random effects. The individuals nested within pedigrees were modeled using a generalized linear mixed model (26–28). In the model, SNP genotypes (categorical variables with three levels) were treated as fixed effects, and the dependencies among members within each family were treated as random effects. As pretreatment TG is the best predictor of response to fenofibrate, pretreatment TG level was included as a covariate in all analyses. We adjusted the multivariate growth curve models for potential confounders, including sex, field center, smoking status, diabetes, age, waist circumference, and the baseline TG.

We used growth parameters (slopes) estimated in a growth curve mixed model to characterize TG changes (24–26, 28) because the growth slopes demonstrated higher heritability than the pre- and post-interventional changes. Thus, we calculated the individual slope of the TG change, adjusted using information from all participants, as the phenotype between time points to assess changes in TG and to test gene-treatment interactions. These results were compared with results using the absolute change in TG level as the outcome of interest, and associations were similar between the two methods. Here we report the growth curve results due to higher heritability estimates than the absolute change and better model fit assessed by Akaike's Information Criterion (AIC). A log transformation of covariates was performed to obtain normal distributions and meet model assumptions. In all models, the only significant covariate was the baseline triglyceride level, $P < 0.0001$.

We used HAPLOVIEW version 3.32 (29) to examine *APOB* SNPs. Estimation of SNP allele frequencies used the maximum-likelihood method. We assessed genotype deviations from Hardy-Weinberg equilibrium (HWE) for each *APOB* SNP using the χ^2 test. We excluded one *APOB* SNP (rs679899) that was not in HWE. We calculated the statistical correlation, r^2 , between two SNPs, from HAPLOVIEW.

We initially considered all P values < 0.05 as nominally significant but further assessed their significance using the conservative Bonferroni correction to account for multiple comparisons. Only significant results after Bonferroni correction are the focus of this report.

RESULTS

Sample description

Table 1 displays demographic and clinical characteristics of the study sample. Overall, the sample contained more women than men. Most covariates were evenly distributed among men and women with the exception of diabetes (more women were diabetic). While body mass index (BMI) was similar between men and women, the waist-to-hip ratio was larger among men.

Table 2 demonstrates the difference in serum lipid values between men and women pre- and post-fenofibrate treatment. TG and LDL-C levels were lower after fenofibrate treatment, while HDL-C levels increased significantly

TABLE 1. Demographic and social characteristics of study sample^a

Characteristic	Men ^b	Women ^c
	N = 453 (47.3%)	N = 505 (52.7%)
Age (in years)	48.8 (16.5)	48.1 (16.3)
Current smoker	34 (7.6%)	39 (7.9%)
Currently consumes alcohol	211 (47.2%)	242 (48.7%)
Diabetic ^d	29 (6.5%)	48 (9.7%)
Education		
1 or more years of high school	125 (28.0%)	144 (29.0%)
1 or more years of trade school	52 (11.6%)	42 (8.5%)
1 or more years of college	213 (47.7%)	274 (55.1%)
1 or more years post-college	57 (12.8%)	37 (7.4%)
Body mass index (kg/m ²)	28.4 (4.9)	28.2 (6.3)
Waist-to-hip ratio	0.95 (0.08)	0.85 (0.08)

^aContinuous characteristics report the mean (standard deviation), and categorical characteristics report the number (%).

^bAmong men, six individuals were missing information for smoking status, alcohol consumption, and educational level. Seven individuals were missing information for diabetes status.

^cAmong women, eight individuals were missing information for smoking status, alcohol consumption, diabetes status, and educational level.

^dSubjects were classified as having type 2 diabetes when fasting plasma glucose concentration was 126 mg/dl or use of insulin or diabetes medication was reported.

among men, but not among women, after fenofibrate treatment. For LDL-C, women responded better to treatment than men, as they had higher pre-fenofibrate LDL-C measurements and lower LDL-C measurements post-fenofibrate than men ($P = 0.0353$).

APOB SNP genotyping and characteristics

Table 3 displays the attributes of the five SNPs in the *APOB* gene, including the accession number (rs #) used by dbSNP, and the justification for choosing each SNP. Three of the four SNPs are located in exons, and all polymorphisms were fairly common, as the minor allele frequencies were all greater than 15%. Only one SNP, rs679899, was not in HWE and was removed from all further analyses. **Table 4** shows pair-wise linkage disequilibrium (LD) statistical correlations, r^2 , between the four SNPs. The r^2 values indicate that some of these SNPs were in modest linkage disequilibrium, with r^2 values between .058 and .371. All of the SNPs changed the amino acid sequence in exons 14, 26, or 29, or were in the *APOB* promoter region. The allelic distribution of each SNP, provided in **Table 5**, demonstrates that 10 of the 12 possible genotypes are common (i.e., frequencies greater than 5%). **Table 5** also shows the geometric mean TG level, pre- and post-fenofibrate treatment by genotype.

TABLE 2. Characteristics of unadjusted blood lipids in the study sample

Measure	Men		Women	
	Pre-Fenofibrate	Post-Fenofibrate	Pre-Fenofibrate	Post-Fenofibrate
	mean (SD)	mean (SD)	mean (SD)	mean (SD)
Triglycerides (mg/dl)	145 (111)	94 (52)	137 (92)	91 (58)
Total cholesterol (mg/dl)	190 (40)	169 (36)	189 (38)	165 (35)
HDL-C (mg/dl)	45 (12)	47 (13)	49 (13)	51 (13)
LDL-C (mg/dl)	122 (30)	108 (32)	120 (31)	101 (31)

TABLE 3. Description of the *APOB* SNPs evaluated

SNP	dbSNP ID ^a	Location	Minor Allele	Minor Allele Frequency	HWE <i>P</i> ^b	Justification
-516 C→T	rs934197	promoter	T	0.334	0.2575	(20–24)
Val591Ala	rs679899 ^c	exon 14	A	0.463	0.0071	(19)
<i>Xba</i> I	rs693	exon 26	G	0.489	0.2708	(20–23)
Pro2739Leu	rs676210	exon 26	T	0.217	0.5386	(19, 20–23, HapMap ^d)
<i>Eco</i> RI (Glu4514Lys)	rs1042031	exon 29	T	0.173	0.7080	(19)

APOB, apolipoprotein B gene; HWE, Hardy-Weinberg equilibrium; SNP, single nucleotide polymorphism.

^aSNP ID in the database dbSNP (www.ncbi.nlm.nih.gov/SNP).

^bHardy-Weinberg equilibrium test.

^cThis SNP was excluded from further analysis.

^dHapMap SNP, early phase I.

APOB SNP association analysis

Table 5 summarizes the single SNP association analyses examining the response to fenofibrate treatment. Fenofibrate treatment did not demonstrate any significant associations for LDL-C or HDL-C levels with any of the *APOB* SNPs assessed (supplementary Tables I and II); however, there was a significant treatment effect for one *APOB* SNP (rs676210) with TG levels. Overall, fenofibrate decreased unadjusted TG measurement by 49 mg/dl (a 35% reduction). For the rs676210 SNP, individuals homozygous for the variant allele (T) demonstrated larger reductions in adjusted TG measurements than those with the CC or CT genotypes.

On the basis of the association of rs676210 with TG response to fenofibrate, we assessed whether demographic or clinical covariates were significantly different by genotype that would explain the association (Table 6). While the majority of clinical variables were not statistically different by rs676210 genotype, waist circumference and waist-to-hip ratio were larger in the TT genotype group, indicating that other components of the metabolic syndrome that are highly correlated with triglyceride levels may mediate the observed finding. However, our results (see Table 5) were adjusted for waist circumference. We performed additional analyses further adjusted for waist-to-hip ratio, but the results remained unchanged (supplementary Table III). Finally, the Minnesota field center yielded 30 of the 46 subjects with the rs676210 TT genotype.

DISCUSSION

To our knowledge, the GOLDN Study is one of the largest open-label fenofibrate trials to-date in a general population, and this analysis is the first to examine the

TABLE 4. Calculated linkage disequilibrium (r^2) between *APOB* SNPs in the study population

	rs934197	rs693	rs676210	rs1042031
rs934197				
rs693	0.371			
rs676210	0.093	0.29		
rs1042031	0.094	0.219	0.058	

SNP, single nucleotide polymorphism.

role of *APOB* polymorphisms in response to fenofibrate treatment. Fenofibrate treatment reduces the production of *APOB* and secretion of VLDL (7, 30). It is likely that genetic variation within the *APOB* locus explains variable triglyceride lowering with fenofibrate. We examined five SNPs in the *APOB* gene, three that were previously associated with plasma lipid levels in various contexts (21, 31, 32) and two additional *APOB* SNPs. One of these SNPs (rs679899) was excluded from our analysis because it was not in HWE. We found a significant association between SNP rs676210 (Pro2739Leu) and TG response to fenofibrate treatment: those homozygous for the rare allele (T) showed a greater TG lowering than did individuals who carried the wild-type allele. These results support the conclusion that *APOB* gene variants modulate the response of TG concentrations to fenofibrate treatment.

Fenofibrate, a synthetic agonist of PPAR α , is one drug in the class of hypolipidemic medications known as fibrates. Fibrates help normalize the dyslipidemia associated with increased triglyceride levels by inducing the lipolysis of triglyceride-rich lipoproteins. The mechanism involves increasing lipoprotein lipase (LPL) gene expression (33) and repressing *APOC3* gene expression (34). Fibrates also lower the LDL-cholesterol level by increasing the rate of receptor-mediated removal of LDL particles by favoring the production of larger, more buoyant particles that bind more readily to LDL receptors (35, 36).

Currently, it is unclear how the genetic variation at the rs676210 *APOB* locus, or an allele in LD, might enhance the triglyceride-lowering effect of fenofibrate. In this study, the TT genotype at this SNP site (Pro2739Leu) was associated with a larger response. A leucine residue at amino acid residue 2739 of *APOB* is predicted to be damaging to the structure of the *APOB*100 polypeptide, as judged by SIFT (37) and PolyPhen (38) (supplementary Table IV), and as such, it could reduce either the numbers of VLDL particles assembled, their triglyceride contents, their surface properties, or any combination of these three. Potentially, the VLDL particles made with an *APOB*100 polypeptide containing a leucine rather than a proline residue at position 2739 may have reduced affinity for the LPL inhibitor, *APOC*-III, resulting in the production of fewer, small-dense, triglyceride-rich, LDL-particles that are less

TABLE 5. Triglyceride response to fenofibrate by genotype adjusted for pretreatment triglycerides and other covariates

SNP	Genotype	Number (%)	Pre-Fenofibrate		Post-Fenofibrate		Change in Triglyceride (%)		Change in Triglyceride (mg/dl)	
			Geometric Mean (95% CI)	P ^a	Geometric Mean (95% CI)	P ^a	% Change (95% CI)	P ^a	Absolute Change (95% CI)	P ^a
rs934197	CC	413 (43.1)	113.3 (111.3–115.3)	0.8320	81.3 (79.3–83.4)	0.9659	–25.8 (–21.7 to –30.0)	0.7088	31.8 (29.7–33.9)	0.7431
	CT	440 (45.9)	114.2 (112.2–116.2)		81.0 (79.0–83.0)		–27.0 (–22.9 to –31.1)		30.5 (28.4–32.5)	
	TT	105 (11.0)	112.7 (110.7–114.7)		81.9 (79.8–83.9)		–25.3 (–19.6 to –31.0)		30.9 (28.7–33.0)	
		252 (26.3)	110.7 (108.7–112.7)	0.1524	81.0 (79.0–83.0)	0.9531	–23.7 (–19.3 to –28.2)	0.0847	30.4 (28.3–32.6)	0.8263
rs693	AA	486 (50.7)	114.5 (112.5–116.5)		81.1 (79.0–83.1)		–27.9 (–23.8 to –32.0)		31.6 (29.5–33.7)	
	AG	220 (23.0)	115.2 (113.2–117.2)		81.8 (79.8–83.8)		–26.3 (–21.7 to –30.9)		30.7 (28.6–32.9)	
	GG	593 (61.9)	113.2 (111.2–115.2)	0.7946	82.4 (80.4–84.4)	0.0467	–24.7 (–20.8 to –28.5)	0.0053	29.2 (27.1–31.2)	<0.0001
		319 (33.3)	114.3 (112.3–116.3)		81.0 (78.9–83.0)		–28.3 (–23.9 to –32.6)		33.0 (30.9–35.1)	
rs676210	CT	46 (4.8)	115.3 (113.3–117.4)		71.2 (69.1–73.3)		–34.5 (–27.1 to –41.8)		43.3 (41.2–45.5)	
	CC	654 (68.3)	113.4 (111.4–115.4)	0.8897	80.8 (78.8–82.8)	0.6920	–26.8 (–22.9 to –30.8)	0.6716	32.5 (30.4–34.6)	0.1375
	CT	277 (28.9)	114.3 (112.3–116.3)		82.7 (80.6–84.7)		–25.4 (–20.9 to –29.8)		29.5 (27.4–31.6)	
	TT	27 (2.8)	112.5 (110.4–114.5)		78.8 (76.7–81.0)		–25.2 (–14.9 to –35.5)		24.3 (21.9–26.7)	

All values are adjusted for sex, field center, smoking status, diabetes status, age, waist circumference, baseline triglyceride, and family relatedness with the only significant covariate being baseline triglyceride ($P < 0.0001$).

^aAdditive model.

Supplemental Material can be found at:
<http://www.jlr.org/content/suppl/2010/08/19/jlr.P001834.DC1.html>

rapidly cleared from the circulation than larger, more buoyant LDL. This seems plausible as previous work has shown that APOC-III plays a role in regulating the LPL-mediated lipolysis of APOB-containing lipoproteins (39, 40) and their internalization via LDL receptors (39, 41). It is also possible that the observed association between the rare allele at the rs676210 *APOB* locus and the increased triglyceride-lowering effects of fenofibrate may be mediated through the increased catabolism of all APOB100-containing lipoproteins (7, 10, 42, 43) via increased activity of LPL that occurs in response to this fibrates. Additionally, altered metabolism of APOA1-containing lipoproteins may be involved. Using the protein-protein interactions contained in the STRING (44) database, APOB and PPAR α have a predicted connection through interacting pathways (Fig. 1), where the key molecule tying the pathways together is APOA1, and fibrates are known to induce transcription of *APOA1* (30).

The rs676210 SNP may not be the causative SNP responsible for the demonstrated association: it is in modest linkage disequilibrium with another *APOB* SNP, rs693. However, the rs693 SNP is a synonymous variant that causes no change in the amino acid sequence (45, 46), rendering it an unlikely causal candidate (42, 47). SIFT (37) and PolyPhen (38) suggest that rs676210 is damaging to APOB protein structure, thereby adding confidence to the demonstrated association between TG-lowering and rs676210.

This study has several strengths. Prior work in these data demonstrated genetic homogeneity (16) between the two population groups studied (Utah and Minnesota), thus reducing concerns regarding population stratification. Furthermore, in the analysis, we employed a random coefficient regression model to evaluate the change in the slope of lipid response in addition to the absolute change in lipid response. This is a strength because the slope demonstrated a phenotype that was more heritable than did the absolute change in lipid response (24–26, 28); however, the results were similar when assessing absolute change and percent change. The sample size is relatively large. Lastly, we had duplicate measures of triglycerides at each time point (before and after fenofibrate treatment), thereby reducing measurement error.

Despite these strengths, there were also limitations to our study. One of these is absence of concentrations of APOB, limiting our ability to correlate significant polymorphisms in *APOB* with APOB concentration. Furthermore, this study examined an ethnically homogeneous sample—Caucasians of European ancestry—and these results may not generalize to other ethnic populations due to ethnic differences in allele frequencies and patterns of linkage disequilibrium. The *APOB* gene is fairly large, yet we only examined four SNPs in the gene. While the density of these SNPs throughout the *APOB* gene is of concern, the ones we selected were almost exclusively from exons. Although the rs676210 is not a tag SNP for the Caucasian population, the SNP is in a block and its haplotype is tagged by rs1042034, located at 21,078,786 and covering

TABLE 6. Demographic variable distribution based on rs676210 genotype

Characteristic	CC (n = 593)	CT (n = 319)	TT (n = 46)	P
Sex (male)	281 (47.4%)	144 (45.1%)	28 (60.9%)	0.1356
Field center (Minnesota)	266 (44.9%)	166 (52.0%)	30 (65.2%)	0.0072
Current smoker	49 (8.4%)	20 (6.3%)	4 (8.7%)	0.5177
Currently consumes alcohol	277 (47.6%)	149 (47.2%)	27 (58.7%)	0.3267
Diabetic	48 (8.3%)	23 (7.3%)	6 (13.0%)	0.4069
Age (in years)	48.2 (15.9)	48.1 (16.9)	54.8 (15.5)	0.0785
Education				0.6608
1 or more years of high school	168 (28.9%)	87 (27.5%)	14 (30.4%)	
1 or more years of trade school	55 (9.5%)	35 (11.1%)	4 (8.7%)	
1 or more years of college	303 (52.1%)	164 (51.9%)	20 (43.5%)	
1 or more years post-college	56 (9.6%)	30 (9.5%)	8 (17.4%)	
Body mass index (kg/m ²)	28.4 (5.6)	28.0 (5.8)	30.0 (6.4)	0.0517
Waist circumference (cm)	96.6 (15.9)	95.0 (15.8)	103.4 (21.8)	0.0014
Waist-to-hip ratio	0.89 (0.09)	0.89 (0.09)	0.95 (0.2)	0.0003

an area of 20,054 bp. Lastly, the association between rs676210 and triglyceride response has not yet been replicated.

In summary, these findings support a role for *APOB* gene variants in predicting TG response to fenofibrate treatment that warrants further analyses in additional

patient/population samples. In this time of the burgeoning possibilities of personalized medicine, if this association is demonstrated in other studies, *APOB* polymorphisms, specifically the rs676210 variant, may aid in identifying individuals who will benefit the most from the TG-lowering effects of fenofibrate. **Fig 1**

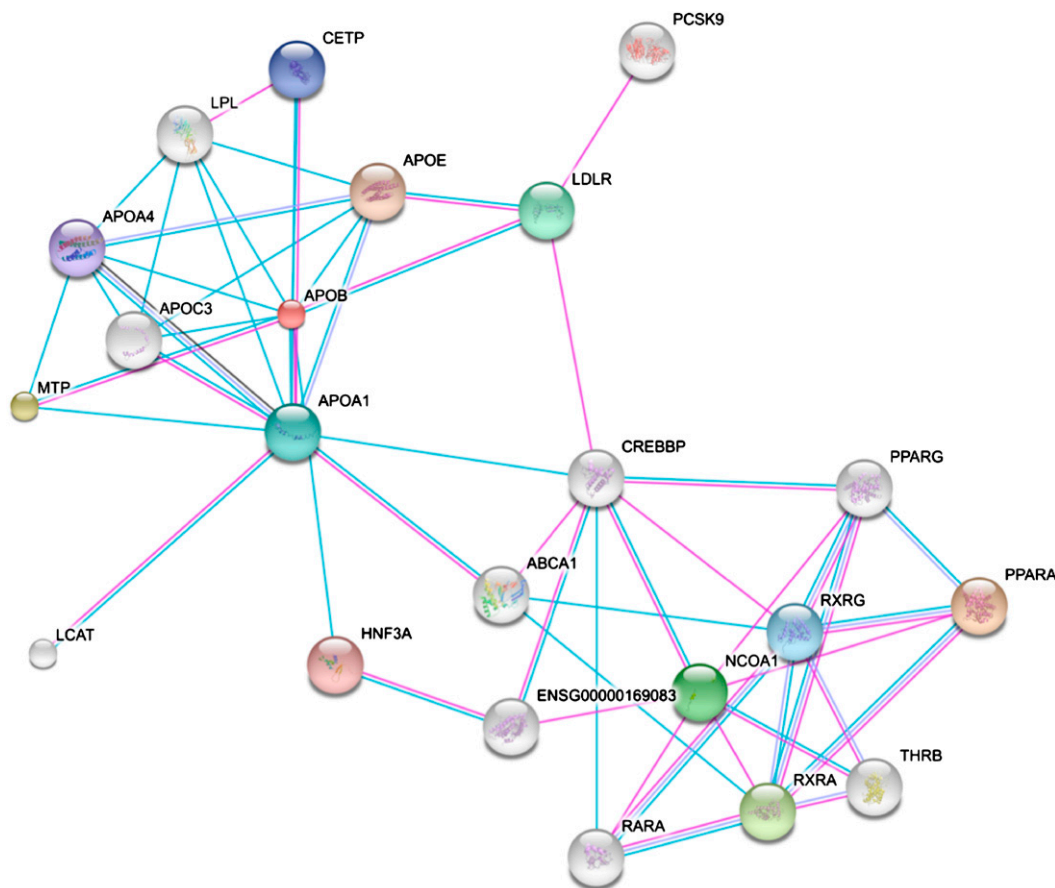


Fig. 1. Evidence-based diagram of known protein-protein interactions between PPAR α and APOB from the STRING database. Pink lines demonstrate experimental evidence, blue lines are database evidence, and purple lines are homology evidence. APO, apolipoprotein; CETP, cholesterylester transfer protein; CREBBP, CREB-binding protein; HNF, hepatocyte nuclear factor; LCAT, lecithin-cholesterol acyltransferase; LDLR, low density lipoprotein receptor; MTP, microsomal triglyceride transfer protein; NCOA1, nuclear receptor coactivator 1; PCSK9, proprotein convertase subtilisin/kexin 9; PPAR, peroxisome proliferator activated receptor; RAR, retinoic acid receptor; RXR, retinoid X receptor; THR β , thyroid hormone receptor beta.

The authors would like to thank the families for their participation in this research. We wish to acknowledge Abbott Laboratories (Abbott Park, IL) for its supply of study medication for this project.

REFERENCES

- Sarwar, N., J. Danesh, G. Eiriksdottir, G. Sigurdsson, N. Wareham, S. Bingham, S. M. Boekholdt, K. T. Khaw, and V. Gudnason. 2007. Triglycerides and the risk of coronary heart disease: 10,158 incident cases among 262,525 participants in 29 Western prospective studies. *Circulation*. **115**: 450–458.
- Chiodini, B. D., S. Barlera, M. G. Franzosi, V. L. Beceiro, M. Inrona, and G. Tognoni. 2003. APO B gene polymorphisms and coronary artery disease: a meta-analysis. *Atherosclerosis*. **167**: 355–366.
- Ozturk, I. C., and A. A. Killeen. 1999. An overview of genetic factors influencing plasma lipid levels and coronary artery disease risk. *Arch. Pathol. Lab. Med.* **123**: 1219–1222.
- Sofi, F., A. M. Gori, R. Marcucci, G. Innocenti, C. Dini, S. Genise, G. F. Gensini, R. Abbate, C. Surrenti, and A. Casini. 2007. Adherence to a healthful life attenuates lipid parameters among a healthy Italian population. *Nutr. Metab. Cardiovasc. Dis.* **17**: 642–648.
- Ford, E. S., C. Li, G. Zhao, W. S. Pearson, and A. H. Mokdad. 2009. Hypertriglyceridemia and its pharmacologic treatment among US adults. *Arch. Intern. Med.* **169**: 572–578.
- Robins, S. J., and H. E. Bloomfield. 2006. Fibrin acid derivatives in cardiovascular disease prevention: results from the large clinical trials. *Curr. Opin. Lipidol.* **17**: 431–439.
- Keating, G. M., and K. F. Croom. 2007. Fenofibrate: a review of its use in primary dyslipidaemia, the metabolic syndrome and type 2 diabetes mellitus. *Drugs*. **67**: 121–153.
- Zaman, M. M., S. Ikemoto, N. Yoshiike, C. Date, T. Yokoyama, and H. Tanaka. 1997. Association of apolipoprotein genetic polymorphisms with plasma cholesterol in a Japanese rural population. The Shibata Study. *Arterioscler. Thromb. Vasc. Biol.* **17**: 3495–3504.
- Knott, T. J., S. C. Rall, Jr., T. L. Innerarity, S. F. Jacobson, M. S. Urdea, B. Levy-Wilson, L. M. Powell, R. J. Pease, R. Eddy, H. Nakai, et al. 1985. Human apolipoprotein B: structure of carboxyl-terminal domains, sites of gene expression, and chromosomal localization. *Science*. **230**: 37–43.
- Fisher, W. R., L. A. Zech, and P. W. Stacpoole. 1994. ApoB metabolism in familial hypercholesterolemia. Inconsistencies with the LDL receptor paradigm. *Arterioscler. Thromb.* **14**: 501–510.
- Olofsson, S. O., L. Asp, and J. Boren. 1999. The assembly and secretion of apolipoprotein B-containing lipoproteins. *Curr. Opin. Lipidol.* **10**: 341–346.
- Brodsky, J. L., and E. A. Fisher. 2008. The many intersecting pathways underlying apolipoprotein B secretion and degradation. *Trends Endocrinol. Metab.* **19**: 254–259.
- Rajput-Williams, J., T. J. Knott, S. C. Wallis, P. Sweetnam, J. Yarnell, N. Cox, G. I. Bell, N. E. Miller, and J. Scott. 1988. Variation of apolipoprotein-B gene is associated with obesity, high blood cholesterol levels, and increased risk of coronary heart disease. *Lancet*. **2**: 1442–1446.
- Marshall, H. W., L. C. Morrison, L. L. Wu, J. L. Anderson, P. S. Corneli, D. M. Stauffer, A. Allen, L. A. Karagounis, and R. H. Ward. 1994. Apolipoprotein polymorphisms fail to define risk of coronary artery disease. Results of a prospective, angiographically controlled study. *Circulation*. **89**: 567–577.
- Higgins, M., M. Province, G. Heiss, J. Eckfeldt, R. C. Ellison, A. R. Folsom, D. C. Rao, J. M. Sprafka, and R. Williams. 1996. NHLBI Family Heart Study: objectives and design. *Am. J. Epidemiol.* **143**: 1219–1228.
- Pankow, J. S., M. A. Province, S. C. Hunt, and D. K. Arnett. 2002. Regarding “Testing for population subdivision and association in four case-control studies”. *Am. J. Hum. Genet.* **71**: 1478–1480.
- Livak, K. J. 1999. Allelic discrimination using fluorogenic probes and the 5′ nuclease assay. *Genet. Anal.* **14**: 143–149.
- Bentzen, J., P. Poulsen, A. Vaag, and M. Fenger. 2003. Further studies of the influence of apolipoprotein B alleles on glucose and lipid metabolism. *Hum. Biol.* **75**: 687–703.
- Perez-Martinez, P., F. Perez-Jimenez, J. M. Ordovas, C. Bellido, J. A. Moreno, P. Gomez, C. Marin, R. A. Fernandez de la Puebla, J. A. Paniagua, and J. Lopez-Miranda. 2007. The APOB -516C/T polymorphism has no effect on lipid and apolipoprotein response following changes in dietary fat intake in a healthy population. *Nutr. Metab. Cardiovasc. Dis.* **17**: 224–229.
- Perez-Martinez, P., F. Perez-Jimenez, J. M. Ordovas, J. A. Moreno, C. Marin, R. Moreno, Y. Jimenez-Gomez, J. A. Paniagua, and J. Lopez-Miranda. 2007. Postprandial lipemia is modified by the presence of the APOB-516C/T polymorphism in a healthy Caucasian population. *Lipids*. **42**: 143–150.
- Lopez-Miranda, J., J. M. Ordovas, M. A. Ostos, C. Marin, S. Jansen, J. Salas, A. Blanco-Molina, J. A. Jimenez-Perez, F. Lopez-Segura, and F. Perez-Jimenez. 1997. Dietary fat clearance in normal subjects is modulated by genetic variation at the apolipoprotein B gene locus. *Arterioscler. Thromb. Vasc. Biol.* **17**: 1765–1773.
- Lopez-Miranda, J., C. Marin, P. Castro, P. Gomez, A. Gonzalez-Amieva, E. Paz, D. Bravo, J. M. Ordovas, J. Jimenez-Perez, and F. Perez-Jimenez. 2000. The effect of apolipoprotein B xbaI polymorphism on plasma lipid response to dietary fat. *Eur. J. Clin. Invest.* **30**: 678–684.
- Matys, V., E. Fricke, R. Geffers, E. Gossling, M. Haubrock, R. Hehl, K. Hornischer, D. Karas, A. E. Kel, O. V. Kel-Margoulis, et al. 2003. TRANSFAC: transcriptional regulation, from patterns to profiles. *Nucleic Acids Res.* **31**: 374–378.
- Corbett, J., A. Kraja, I. B. Borecki, and M. A. Province. 2003. Use of a random coefficient regression (RCR) model to estimate growth parameters. *BMC Genet.* **4**(Suppl. 1): S5.
- Laird, N. M., and J. H. Ware. 1982. Random-effects models for longitudinal data. *Biometrics*. **38**: 963–974.
- Littell, R. C., G. A. Milliken, W. W. Stroup, and R. D. Wolfinger. 1996. SAS System for Mixed Models. SAS Institute, Inc., Cary, NC.
- Singer, J. D. 1998. Using SAS PROC MIXED to fit multilevel models, hierarchical models, and individual growth models. *J. Educ. Behav. Stat.* **24**: 323–355.
- Lai, C. Q., S. Demissie, L. A. Cupples, Y. Zhu, X. Adiconis, L. D. Parnell, D. Corella, and J. M. Ordovas. 2004. Influence of the APOA5 locus on plasma triglyceride, lipoprotein subclasses, and CVD risk in the Framingham Heart Study. *J. Lipid Res.* **45**: 2096–2105.
- Barrett, J. C., B. Fry, J. Maller, and M. J. Daly. 2005. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics*. **21**: 263–265.
- Staels, B., J. Dallongeville, J. Auwerx, K. Schoonjans, E. Leitersdorf, and J. C. Fruchart. 1998. Mechanism of action of fibrates on lipid and lipoprotein metabolism. *Circulation*. **98**: 2088–2093.
- Gylling, H., K. Kontula, U. M. Koivisto, H. E. Miettinen, and T. A. Miettinen. 1997. Polymorphisms of the genes encoding apoproteins A-I, B, C-III, and E and LDL receptor, and cholesterol and LDL metabolism during increased cholesterol intake. Common alleles of the apoprotein E gene show the greatest regulatory impact. *Arterioscler. Thromb. Vasc. Biol.* **17**: 38–44.
- Jemaa, R., A. Mebazaa, and F. Fumeron. 2004. Apolipoprotein B signal peptide polymorphism and plasma LDL-cholesterol response to low-calorie diet. *Int. J. Obes. Relat. Metab. Disord.* **28**: 902–905.
- Heller, F., and C. Harvengt. 1983. Effects of clofibrate, bezafibrate, fenofibrate and probucol on plasma lipolytic enzymes in normolipemic subjects. *Eur. J. Clin. Pharmacol.* **25**: 57–63.
- Staels, B., N. Vu-Dac, V. A. Kosykh, R. Saladin, J. C. Fruchart, J. Dallongeville, and J. Auwerx. 1995. Fibrates downregulate apolipoprotein C-III expression independent of induction of peroxisomal acyl coenzyme A oxidase. A potential mechanism for the hypolipidemic action of fibrates. *J. Clin. Invest.* **95**: 705–712.
- Caslake, M. J., C. J. Packard, A. Gaw, E. Murray, B. A. Griffin, B. D. Vallance, and J. Shepherd. 1993. Fenofibrate and LDL metabolic heterogeneity in hypercholesterolemia. *Arterioscler. Thromb.* **13**: 702–711.
- Guerin, M., E. Bruckert, P. J. Dolphin, G. Turpin, and M. J. Chapman. 1996. Fenofibrate reduces plasma cholesteryl ester transfer from HDL to VLDL and normalizes the atherogenic, dense LDL profile in combined hyperlipidemia. *Arterioscler. Thromb. Vasc. Biol.* **16**: 763–772.
- Ng, P. C., and S. Henikoff. 2003. SIFT: predicting amino acid changes that affect protein function. *Nucleic Acids Res.* **31**: 3812–3814.
- Sunyaev, S., V. Ramensky, I. Koch, W. Lathe 3rd, A. S. Kondrashov, and P. Bork. 2001. Prediction of deleterious human alleles. *Hum. Mol. Genet.* **10**: 591–597.
- Windler, E., and R. J. Havel. 1985. Inhibitory effects of C apolipoproteins from rats and humans on the uptake of triglyceride-rich

- lipoproteins and their remnants by the perfused rat liver. *J. Lipid Res.* **26**: 556–565.
40. Wang, C. S., W. J. McConathy, H. U. Kloer, and P. Alaupovic. 1985. Modulation of lipoprotein lipase activity by apolipoproteins. Effect of apolipoprotein C-III. *J. Clin. Invest.* **75**: 384–390.
41. Beisiegel, U., W. Weber, and G. Bengtsson-Olivecrona. 1991. Lipoprotein lipase enhances the binding of chylomicrons to low density lipoprotein receptor-related protein. *Proc. Natl. Acad. Sci. USA.* **88**: 8342–8346.
42. Aalto-Setälä, K., K. Kontula, M. Manttari, J. Huttunen, V. Manninen, P. Koskinen, and H. M. Frick. 1991. DNA polymorphisms of apolipoprotein B and AI/CIII genes and response to gemfibrozil treatment. *Clin. Pharmacol. Ther.* **50**: 208–214.
43. Watts, G. F., P. H. Barrett, J. Ji, A. P. Serone, D. C. Chan, K. D. Croft, F. Loehrer, and A. G. Johnson. 2003. Differential regulation of lipoprotein kinetics by atorvastatin and fenofibrate in subjects with the metabolic syndrome. *Diabetes.* **52**: 803–811.
44. Jensen, L. J., M. Kuhn, M. Stark, S. Chaffron, C. Creevey, J. Muller, T. Doerks, P. Julien, A. Roth, M. Simonovic, et al. 2009. STRING 8—a global view on proteins and their functional interactions in 630 organisms. *Nucleic Acids Res.* **37**: D412–D416.
45. Islam, M. S., O. T. Raitakari, M. Juonala, M. Kahonen, N. Hutri-Kahonen, A. Collings, K. Aalto-Setälä, K. Kontula, J. Marniemi, J. S. Viikari, et al. 2005. Apolipoprotein A-I/C-III/A-IV SstI and apolipoprotein B XbaI polymorphisms and their association with carotid artery intima-media thickness in the Finnish population. The Cardiovascular Risk in Young Finns Study. *Atherosclerosis.* **180**: 79–86.
46. Scartezini, M., M. A. Zago, E. A. Chautard-Freire-Maia, A. Pazin-Filho, J. A. Marin-Neto, J. K. Hotta, A. J. Nascimento, and J. E. Dos-Santos. 2003. The X–X-/E+E+ genotype of the XbaI/EcoRI polymorphisms of the apolipoprotein B gene as a marker of coronary artery disease in a Brazilian sample. *Braz. J. Med. Biol. Res.* **36**: 369–375.
47. Misra, A., S. Nishanth, S. T. Pasha, R. M. Pandey, P. Sethi, and D. S. Rawat. 2001. Relationship of XbaI and EcoRI polymorphisms of apolipoprotein-B gene to dyslipidemia and obesity in Asian Indians in North India. *Indian Heart J.* **53**: 177–183.